

This article was downloaded by:

On: 26 January 2011

Access details: *Access Details: Free Access*

Publisher *Taylor & Francis*

Informa Ltd Registered in England and Wales Registered Number: 1072954 Registered office: Mortimer House, 37-41 Mortimer Street, London W1T 3JH, UK



## Nucleosides, Nucleotides and Nucleic Acids

Publication details, including instructions for authors and subscription information:

<http://www.informaworld.com/smpp/title~content=t713597286>

### Novel Types of N<sup>6</sup>,2'-Cyclonucleosides

Jean M. J. Tronchet<sup>a</sup>; Rachid Benhamza<sup>a</sup>; Gérald Bernardinelli<sup>a</sup>

<sup>a</sup> Departments of Pharmaceutical Chemistry and Crystallography, Faculty of Sciences, Geneva 4, Switzerland

**To cite this Article** Tronchet, Jean M. J. , Benhamza, Rachid and Bernardinelli, Gérald(1993) 'Novel Types of N<sup>6</sup>,2'-Cyclonucleosides', *Nucleosides, Nucleotides and Nucleic Acids*, 12: 1, 55 — 71

**To link to this Article:** DOI: 10.1080/07328319308016194

**URL:** <http://dx.doi.org/10.1080/07328319308016194>

PLEASE SCROLL DOWN FOR ARTICLE

Full terms and conditions of use: <http://www.informaworld.com/terms-and-conditions-of-access.pdf>

This article may be used for research, teaching and private study purposes. Any substantial or systematic reproduction, re-distribution, re-selling, loan or sub-licensing, systematic supply or distribution in any form to anyone is expressly forbidden.

The publisher does not give any warranty express or implied or make any representation that the contents will be complete or accurate or up to date. The accuracy of any instructions, formulae and drug doses should be independently verified with primary sources. The publisher shall not be liable for any loss, actions, claims, proceedings, demand or costs or damages whatsoever or howsoever caused arising directly or indirectly in connection with or arising out of the use of this material.

## NOVEL TYPES OF $N^6,2'$ -CYCLONUCLEOSIDES

Jean M. J. Tronchet,\* Rachid Benhamza and Gérald Bernardinelli

Departments of Pharmaceutical Chemistry and Crystallography, Faculty of Sciences, 30 Quai Ernest-Ansermet, 1211 Geneva 4, Switzerland

**Abstract :** Upon oxidation followed by treatment with hydroxylamine, the 3',5'-diblocked uridine **1** gave the expected oxime **2** together with the  $N^6,2'$ -cyclonucleoside **3** formed by nucleophilic attack of hydroxylamine at both C-6 and C-2' positions. Reduction of **2** took place predominantly from the  $\alpha$  face and the major *D-arabino* compound obtained gave the cyclonucleoside **7** *via* Michael type addition. The structures of the novel cyclonucleosides, particularly their configuration at C-6 were established by X-ray diffraction.

Deoxy *N*-hydroxyamino sugars and nucleosides are very close analogs of their oxygenated counterparts and present the interesting property of oxidizing spontaneously in solution to give the corresponding nitroxide free radical in a stationary concentration sufficient to provide good EPR spectra and too small to significantly degrade the resolution of their NMR spectra.<sup>1</sup> We have previously described 3'-deoxy-3'-*N*-hydroxyamino analogs of nucleosides<sup>2</sup> and report here the synthesis and properties of some derivatives of 2'-deoxy-2'-*N*-hydroxyamino uridine. Owing principally to the so-called  $\alpha$ -effect,<sup>3</sup> hydroxylamines are very efficient nucleophiles towards activated C-C double bonds and this property renders feasible a scarcely encountered mode of formation of cyclonucleosides : Michael type reactions.<sup>4</sup> This led to a novel type of cyclonucleosides  $N^6,2'$ -cyclo-5,6-dihydrouridine derivatives. Part of this work has been the subject of a preliminary communication.<sup>5</sup>

TABLE 1.  $^1\text{H}$  NMR Chemical Shifts of Uridine Derivatives (200 MHz)

Compd	H-1'	H-2'	H-3'	H-4'	H-5'	H-5	H-6
<b>2a<sup>a</sup></b>	6.16		5.22	3.84	4.02 4.08	5.69	7.10
<b>2b<sup>a</sup></b>	6.18		5.24	3.90	4.11	5.73	7.12
<b>3<sup>a</sup></b>	5.58		4.73	3.83	4.10	2.85 3.08	4.69
<b>4<sup>b</sup></b>	5.55		4.42	3.77	3.66 3.86	2.75 2.95	4.65
<b>5<sup>a</sup></b>	5.77		5.60	4.22	4.22	3.0	4.95
<b>6a<sup>b</sup></b>	6.55		4.93	3.72- 3.95	3.72- 3.95	5.67	7.57
<b>6b<sup>b</sup></b>	6.42		4.81	3.72- 3.95	3.72- 3.95	5.66	7.52
<b>7<sup>c</sup></b>	5.85	3.83	3.72	3.39	3.42 3.61	2.80 2.91	4.58
<b>9<sup>a</sup></b>	6.20	4.41	5.35	4.07	4.19 4.33	2.93 3.10	5.02
<b>10<sup>b</sup></b> <i>D-arabino</i>	6.38	4.90	5.06	3.96	3.87 3.96	5.59	8.03
<b>10<sup>b</sup></b> <i>D-ribo</i>	6.49	4.90	4.60	4.39	3.82 3.98	5.70	8.02

<sup>a</sup> In  $\text{CDCl}_3$ . <sup>b</sup> In  $\text{CD}_3\text{OD}$ . <sup>c</sup> In  $(\text{CD}_3)_2\text{SO}$ .

The 3',5'-diblocked uridine **1<sup>6</sup>** was found the most suitable substrate for modifying the 2' position. It was oxidized by chromic anhydride following Garegg's procedure<sup>7</sup> to the expected ketonucleoside which was not isolated but directly oximated to a 1:3 mixture of the geometrical isomers (**2a** and **2b**) of **2** (45.5%). The cyclonucleoside **3** (10%) was also obtained. As evident from the NMR data of **2** (TABLES 1-3), a reliable assignment of the *E-Z*

TABLE 2. <sup>1</sup>H NMR Coupling Constants of Uridine Derivatives<sup>a</sup>

Compd	<i>J</i> <sub>1',2'</sub>	<i>J</i> <sub>1',3'</sub>	<i>J</i> <sub>2',3'</sub>	<i>J</i> <sub>3',4'</sub>	<i>J</i> <sub>4',5'</sub>	<i>J</i> <sub>5',5'</sub>	<i>J</i> <sub>5,6</sub>	<i>J</i> <sub>5,5</sub>
<b>2a</b>		1.8		8.0	3.5 5.0	12.0	8.0	
<b>2b</b>		1.8		8.0	4.0 5.0		8.4	
<b>3</b>				8.0	2.7		10.0 5.2	16.5
<b>4</b>				8.7	4.0 1.0	12.0	9.5 5.5	16.2
<b>5</b>				8.0			8.0	
<b>6a</b>		1.5		6.0			8.0	
<b>6b</b>		1.5		7.0			8.0	
<b>7</b>	6.0		6.0	8.5	5.5 5.5	10.5	6.7	17.7
<b>9</b>	6.2		6.0	7.5	5.0 3.2	12.5	8.0 7.0	17.5
<b>10</b> D-arabino	7.0			6.2	? 2.5	14.5	8.0	
<b>10</b> D-ribo	2.0		7.5	8.5	3.0 2.2	14.5	8.0	

<sup>a</sup> Same solvents as in TABLE 1.

configuration was not possible (very close values of  $\delta_{H-1}$  and  $\delta_{H-3}$  of the two isomers in particular). The 5,6-dihydrouracil moiety of compound **3** gave the expected NMR signals and the structure of **3** was further established by its acetylation to an unisolated di-*O*-acetyl [(NMR:  $\delta$  2.06, 2.08, 2s, 2x3H, 2 OAc; MS: 558 (10.3,  $M^+$  - Ac), 542 (3.1,  $M^+$  - AcO)] derivative. Glycosidation at position 2' and de-*O*-silylation of compound **3** using acidic methanol gave **4** (94%) which was acetylated to **5** (SCHEME 1). X-ray diffraction study of

TABLE 3.  $^{13}\text{C}$  NMR Chemical Shifts of Uridine Derivatives (50.4 MHz)<sup>a</sup>

Compd	C-1'	C-2'	C-3'	C-4'	C-5'	C-5	C-6
<b>2a</b>	84.66	155.60	69.20	83.55	63.59	102.76	141.88
<b>2b</b>	82.45	156.88	70.59	82.45	62.23	102.85	142.75
<b>3</b>	89.27	97.96	67.82	82.04	60.48	35.66	67.82
<b>4</b>	87.28	102.37	68.79	84.46	61.76	37.26	72.44
<b>6a</b>	85.49	151.57	69.55	83.36	62.33	102.81	144.89
<b>6b</b>	86.08	152.19	66.96	83.79	62.49	103.21	144.12
<b>7</b>	87.62	71.77	73.92	83.94	61.39	31.26	81.66
<b>9</b>	86.73	72.35	72.35	76.61	62.69	31.67	79.29
<b>10</b> D-arabino			71.40		62.00	101.60	
<b>10</b> D-ribo			70.50		62.00	103.00	

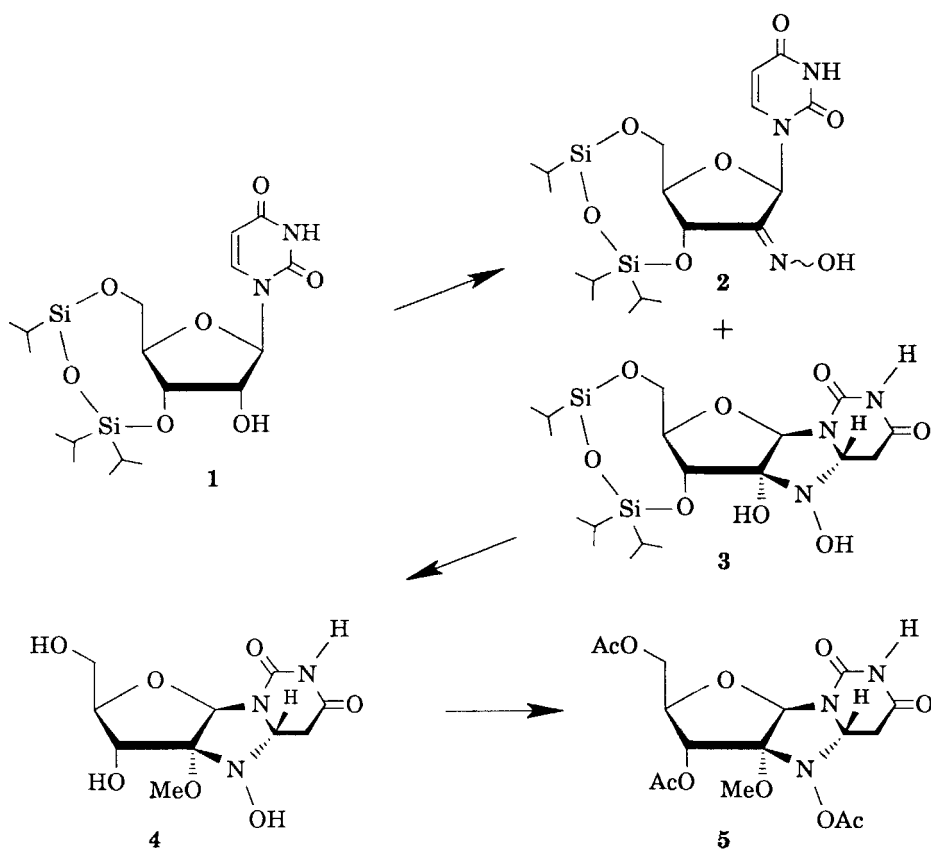
<sup>a</sup> Same solvents as in TABLE 1 except for **7** ( $\text{CD}_3\text{OD} + \text{DMSO}-d_6$ ).

**5** (FIG. 1) showed the configuration at the two new asymmetric carbon atoms to be 6*S*,2'*R*.

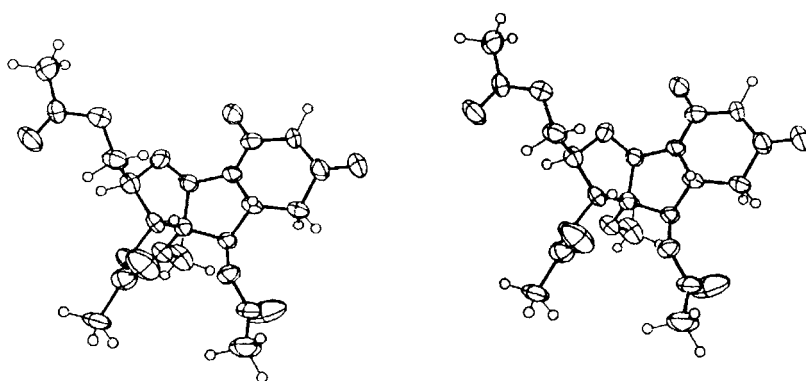
De-O-silylation of **2** gave a 3:7 mixture of geometrical isomers **6** as a hygroscopic solid for which no acceptable elementary analysis could be obtained. Its structure was instead asserted by high resolution mass spectroscopy ( $\text{M}^+ m/z$  calcd : 257.0647, found : 257.0672).

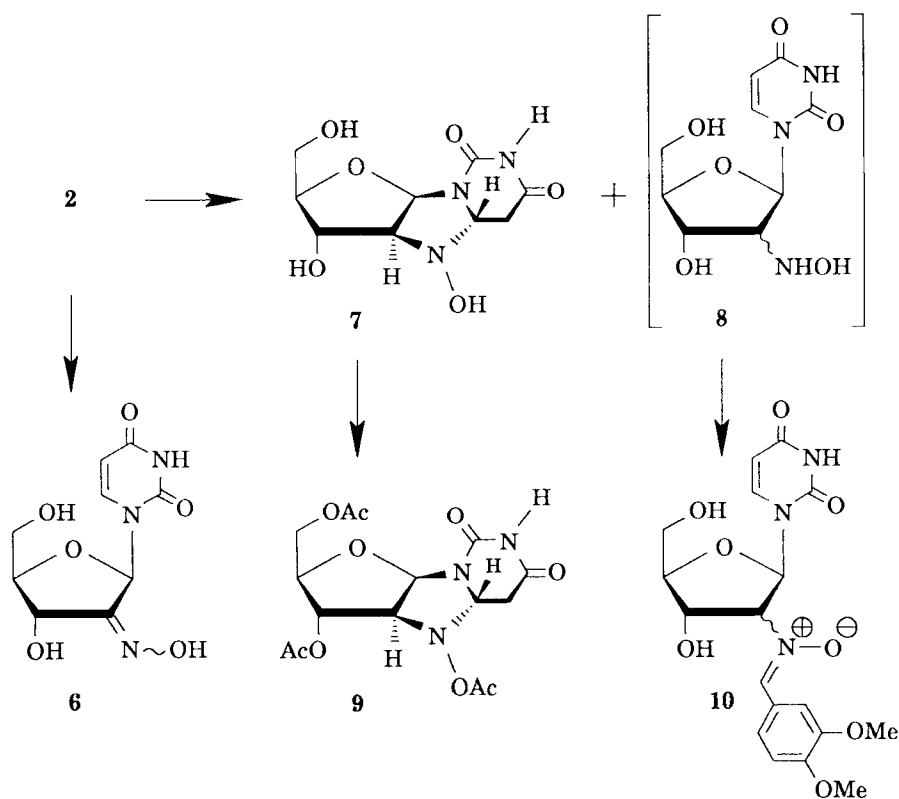
Upon acidic desilylation followed by reduction ( $\text{BH}_3$ -pyridine), **2** gave a mixture, in variable proportions, of the cyclonucleoside **7** and an unresolvable unstable mixture of the epimers **8** (SCHEME 2).

Compound **7** was acetylated to **9** (64%) and **8** converted to the corresponding nitrones **10**. Typically, we did not isolate **7** after the reduction stage but treated the reaction mixture containing **7** and **8** with veratraldehyde to obtain unreacted **7** (75% from **2**) and the unresolvable epimeric mixture of the nitrones **10** (18.5% from **2**) in an 3:7 *arabino/ribo* ratio. The configuration



SCHEME 1

FIG 1. An ORTEP stereoview of cyclonucleoside **5**



SCHEME 2

of each of the epimers **10** was easily assigned from the  $J_{1,2'}$  values (TABLE 2) in particular the low value (2.0 Hz) for the *ribo* isomer establishing the *trans* relationship of H-1' and H-2'. This showed that the reduction proceeded predominantly from the  $\alpha$  side of the furanose ring and that most of the *D-arabino* hydroxylamine **8** underwent conjugate addition onto the uracil moiety leading to **7**. An X-ray diffraction study of **7** established its (6*S*)- $\beta$ -*D-arabino* configuration (FIG. 2).

Besides the configurational assessment they allowed, the X-ray studies provided useful structural information on the novel family of cyclonucleosides represented by **5** and **7**. The furanose ring of **5** adopts a  $^3T_4$  conformation<sup>8</sup> whereas **7** exhibits a quasi perfect  $^3E$  [(C3')-endo] conformation with a minimum value of the asymmetry parameter<sup>9</sup>  $\Delta C_s = 0.005$ . Both

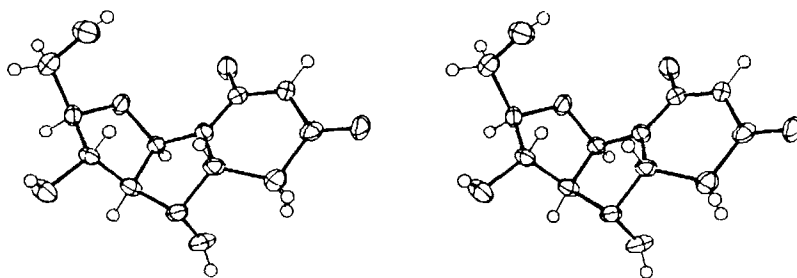


FIG. 2. An ORTEP stereoview of cyclonucleoside 7

furanose rings show maximum puckering amplitude<sup>10</sup> associated with the C(3') atom. In compound 7 the C(4')-C(5') bond adopts the preferred *gauche-gauche* conformation ( $\varphi_{CO} = 62.2(5)^\circ$ ,  $\varphi_{OO} = -55.3(5)^\circ$ ) and the hydroxyl group participate to a bifurcated intermolecular hydrogen bond (TABLE 4). This was not observed in compound 5 where the hydroxyl group was blocked and the C(4')-C(5') bond shows the less favourable *trans-gauche* conformation [ $\varphi_{CO} = 168(1)^\circ$ ,  $\varphi_{OO} = -54(1)^\circ$ ].

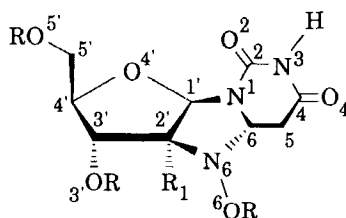
Even if the absolute configuration at the N(6) atom of the imidazolidine rings is *R* for both 5 and 7, their relative configuration differs. In 5, the OAc group is *endo* (pseudo equatorial) relative to the tetrahydro-furanoimidazolidine bicycle and steric interaction of O(6) with H(C(3')) leads N(6) to be *exo* in a  ${}^{C(6)}T_{N(6)}$  conformation of the imidazolidine. In compound 7, the hydroxyl substituent of N(6) is *exo* (pseudo-axial) and the imidazolidine ring adopt a  ${}^{N(6)}T_{C(2')}$  conformation. It is clear that the orientation of the base relative to the sugar moiety, described by the glycosidic torsion angle  $\chi_{CN}$  [O(4')-C(1')-N(1)-C(2)],<sup>11</sup> is fixed by the presence of the nitrogen bridge. In both cases, the glycosidic linkage is restricted to a *syn* arrangement with  $\chi_{CN}$  values of  $-70(2)^\circ$  and  $-62.1(5)^\circ$  for compounds 5 and 7 respectively. The C(1')-N(1) glycosidic bond lengths (1.44 and 1.45 Å) are significantly shorter than the mean value of 1.52 Å observed<sup>12</sup> for compounds where  $\chi_{CN}$  is  $180^\circ$ .

Both dihydrouracil moieties exhibit a half-chair conformation with a minimum value of the asymmetry parameter (TABLE 5) associated with a pseudo  $C_2$  symmetry passing through the C(2)-N(3) bond. In 5, the half-



TABLE 4. Selected Interatomic Distances and Hydrogen Bonds (Å) with e.s.d.'s in parenthesis.

	5	7		5	7
O(4') - C(1')	1.40(1)	1.424(5)	N(1) - C(6)	1.44(1)	1.475(5)
O(4') - C(4')	1.43(1)	1.468(5)	C(2) - O(2)	1.25(1)	1.221(5)
C(1') - C(2')	1.55(1)	1.545(6)	C(2) - N(3)	1.37(2)	1.391(6)
C(1') - N(1)	1.44(2)	1.450(5)	N(3) - C(4)	1.37(2)	1.393(6)
C(2') - C(3')	1.53(2)	1.522(6)	C(4) - O(4)	1.20(2)	1.205(6)
C(2') - N(6)	1.46(1)	1.497(5)	C(4) - C(5)	1.52(1)	1.507(7)
C(3') - C(4')	1.52(2)	1.533(6)	C(5) - C(6)	1.50(1)	1.514(6)
C(3') - O(3')	1.44(1)	1.417(5)	C(6) - N(6)	1.47(2)	1.484(5)
C(4') - C(5')	1.48(2)	1.513(7)	N(6) - O(6)	1.47(1)	1.461(5)
N(1) - C(2)	1.33(1)	1.347(5)			
5	7				
O(H <sub>2</sub> O)....O(4')	x, y, z-1	2.98(1)	O(3')....O(5')	x-1/2, 1/2-y, 1-z	2.703(5)
O(H <sub>2</sub> O)....O(2)	x, y, z-1	2.93(1)	O(5')....N(6)	x+1, y, z	3.015(5)
O(H <sub>2</sub> O)....O(5')	x, y, z-1	3.16(1)	O(6)....O(4')	x-1, y, z	3.021(4)
N(3).....O(5'')	x+1, y, z	2.90(1)	O(6)....O(2)	x-1, y, z	2.808(5)

5 R = Ac, R<sub>1</sub> = OMe7 R = R<sub>1</sub> = H

chair conformation is quasi perfect ( $\Delta C_2 = 0.011$ ) whereas **7** shows a small deformation toward the twist-boat form ( $\Delta C_2 = 0.044$ ). In both compounds the molecular packing is fixed by hydrogen bonds involving all potential donors (TABLE 4).

Diglyme solutions of **3**, **4** and **7** gave, upon spontaneous oxidation in the air, well-resolved EPR spectra. The data corresponding to experiments run at 20 °C are collected in TABLE 6. Increasing the temperature to 40 and 80 °C did not significantly change the aspect of the spectra but at 110°, the radical generated from **7** decomposed. This indicated that the hyperfine couplings measured at 20 °C corresponded already to time-averaged values. All these compounds showed hyperfine couplings with nitrogen (12.9-14

TABLE 5. Selected Torsional Angles( $^\circ$ ), Ring-Puckering Parameters and Minimum Values of Asymmetry Parameters<sup>a</sup>

	5	7
<b>Furanose ring</b>	Conformation ${}^3T_4$	Conformation ${}^3E$
C(4')-O(4')-C(1')-C(2')	-9(1)	-1.2(4)
O(4')-C(1')-C(2')-C(3')	-14(1)	-22.3(4)
C(1')-C(2')-C(3')-C(4')	29(1)	35.3(4)
C(2')-C(3')-C(4')-O(4')	-35(1)	-36.7(4)
C(3')-C(4')-O(4')-C(1')	28(1)	24.1(4)
$Q_2$	0.337	0.374
$\varphi_2$	-60.6	-71.4
$\Delta C_2$	C(1') = 0.019	-
$\Delta C_s$	-	C(3') = 0.005
<b>Dihydrouracil</b>	<i>half-chair</i>	<i>half-chair</i>
C(6)-N(1)-C(2)-N(3)	6(2)	3.6(6)
N(1)-C(2)-N(3)-C(4)	13(2)	23.7(6)
C(2)-N(3)-C(4)-C(5)	0(2)	-11.4(6)
N(3)-C(4)-C(5)-C(6)	-30(2)	-25.2(5)
C(4)-C(5)-C(6)-N(1)	44(2)	46.3(5)
C(5)-C(6)-N(1)-C(2)	-34(2)	-37.9(5)
$Q_T$	0.382	0.443
$\varphi_2$	92.4	99.7
$\theta_2$	117.5	109.2
$\Delta C_2$	(C(2)-N(3)) 0.011	(C(2)-N(3)) 0.044
<b>Imidazolidine ring</b>	Conformation ${}^{C(6)}T_{N(6)}$	Conformation ${}^{N(6)}T_{C(2)}$
N(1)-C(1')-C(2')-N(6)	-16(1)	-25.7(4)
C(1')-C(2')-N(6)-C(6)	35(1)	37.1(4)
C(2')-N(6)-C(6)-N(1)	-40(1)	-33.0(4)
N(6)-C(6)-N(1)-C(1')	30(1)	17.8(4)
C(6)-N(1)-C(1')-C(2')	-9(1)	4.8(4)
$Q_2$	0.371	0.355
$\varphi_2$	83.2	63.2
$\Delta C_2$	C(1') = 0.025	N(1) = 0.034
<b>Miscellaneous</b>		
$\varphi_{CO}$ [C(3')-C(4')-C(5')-O(5')]	168(1)	62.2(5)
$\varphi_{OO}$ [O(4')-C(4')-C(5')-O(5')]	54(1)	-55.3(5)
$\chi_{CN}$ [O(4')-C(1')-N(1)-C(2)]	-70(2)	-62.1(5)
C(3')-C(2')-N(6)-C(6)	-75(1)	-76.6(4)
C(2')-N(6)-C(6)-C(5)	-157(1)	-152.9(4)

<sup>a</sup> For ring-puckering parameters ( $Q_2$ ,  $Q_T$ ,  $\varphi_2$ ,  $\theta_2$ ),<sup>10</sup> the starting position and direction for ring puckering calculations are O(4') to C(4') for furanose, N(1) to C(6) for dihydrouracil and C(1') to N(1) for imidazolidine. Asymmetry parameters ( $\Delta C_s$ ,  $\Delta C_2$ ) are expressed according to Nardelli<sup>9</sup> and conformational nomenclature (E, T,  $\chi_{CN}$ ,  $\varphi_{OO}$ ,  $\varphi_{CO}$ ) based on Sundaralingam's proposals.<sup>8,11</sup>

TABLE 6. EPR Data (diglyme, 20 °C) of Nitroxide Free Radicals from Cyclonucleosides 3, 4 and 7

Compd	g	$a_N$	$a_{H}^\beta$	Extra signal splittings
3	2.0054	13.6	16.4	3x0.5
4	2.0062	12.9	16.0	3x0.5
7	2.0060	14.0	19.3 18.0	0.6, 0.6, 0.5

G). For compounds 3 and 4, one large coupling with a  $\beta$  hydrogen atom was noted. From the modified<sup>13</sup> Rassat's equation<sup>14</sup> [ $a_H = (25 \pm 1) \cos^2\theta$ ] where  $\theta$  stands for the dihedral angle between the C-H bond and the axis of the  $p_z$  orbital on the nitroxide nitrogen atom, this corresponded to an averaged  $\theta$  value of *ca* 38°. As expected from its structure, 7 showed two such  $a_H^\beta$  couplings, both large, this ruling out an envelope conformation of the imidazolidine *N*-oxyl ring in which the nitrogen atom bearing the oxygen would be out of the plane of the four remaining atoms. The measured values of 19.3 and 18.0 G corresponded to  $\theta$  ranges of 26.3-30.5° and 30-33.7° respectively, which indicated a conformation where either C-1' or N-1 (or both) would be out of the general plane of the imidazolidine ring. Some very small long-range hyperfine couplings were also noted but the resolution of the signals did not allow a reliable assignment either to the  $\beta$  nitrogen atom (N-1) or to some of the four  $\gamma$  hydrogen atoms.

Antibacterial activities of compounds 3, 6, 7 and 9 against *E. coli* and *B. subtilis* and activity of 7, 9 and 10 against Polyoma A2 virus have been measured using described procedures.<sup>2</sup> No activity against *E. coli* was found for any of these compounds. A marginal activity against *B. subtilis* was noted for 7 and 9 (minimum inhibitory concentration 5 and 4  $\mu$ M respectively). Concerning the antiviral testing, the only somewhat active compound was the nitrone 10 (partial inhibition of Polyoma A2 virus for a 235  $\mu$ M concentration).

## EXPERIMENTAL

**General Methods.** Melting points (uncorrected) were determined under microscope with a Mettler FP52 melting-point apparatus. Thin layer chromatographies (TLC) were performed on silica gel HF<sub>254</sub> (Merck) with detection by UV light and phosphomolybdic-sulfuric acid.<sup>15</sup> Dry column chromatography<sup>16</sup> was conducted on silica gel 60F<sub>254</sub> (0.063-0.200 mm). Silica gel 60 (0.040-0.200 mm) Merck was used for flash column chromatography.<sup>17</sup> IR spectra were recorded with a Perkin-Elmer Model 357 or a FT-IR Nicolet 20 SXB spectrometers. UV spectra were measured on a Kontron Uvicon 810 spectrophotometer. NMR spectra were recorded at 20 °C on a Bruker WP 200 SY spectrometer (<sup>1</sup>H 200 MHz; <sup>13</sup>C 50.4 MHz; chemical shifts in ppm from TMS;  $\delta$  units). Optical rotations were measured with a Schmidt-Haensch polarimeter and circular dichroism studied with a Jasco S-20 spectropolarimeter. Mass spectra (EIMS, 70 eV) were recorded on a VG-70-70E spectrometer. EPR spectra were recorded on a Varian E-9 spectrometer (X band, 100 KHz modulation) equipped with a variable temperature device. The g values were measured by using a DPPH sample and the magnetic field was calibrated with an NMR marker. All hyperfine coupling constants were checked by simulating the corresponding EPR spectra using a PC program developed in this Laboratory.<sup>18</sup>

**Cristallography.** Experimental data and structure refinement are summarized in TABLE 7. Single crystals were grown at room temperature from H<sub>2</sub>O and EtOH for compound 5 and 7 respectively. Data collection were performed at room temperature on a Nonius CAD4 diffractometer with graphite monochromated Mo K $\alpha$  radiation ( $\lambda = 0.71069$  Å). Diffracted intensities were corrected for Lorentz-polarization but not for absorption. The structures were solved by direct methods (MULTAN-87)<sup>19</sup> and refined by full-matrix least squares with XTAL program.<sup>20</sup> All coordinates of H atoms were calculated for 5 and observed and refined for 7. The polar origin of 5 have been constrained by fixing the y coordinate of O(4'). Crystallographic data have been previously<sup>5</sup> deposited with the Cambridge Crystallographic Data Center, University Chemical Laboratory, Lensfield Road, Cambridge CB21EW, England.

TABLE 7. Summary of Crystal Data, Intensity Measurement and Structure Refinement for Compounds 5 and 7

	5	7
Formula	$C_{16}H_{21}N_3O_{10} \cdot H_2O$	$C_9H_{13}N_3O_6$
Mol. wt.	433.4	259.2
Crystal system	Monoclinic	Orthorhombic
Space Group	$P2_1$	$P2_12_12_1$
a (Å)	10.6418(12)	6.7051(9)
b (Å)	7.4341(7)	10.2824(13)
c (Å)	12.7910(14)	15.593(2)
$\beta$ (°)	96.04(1)	-
V (Å <sup>3</sup> )	1006.3(2)	1075.1(2)
Z	2	4
F(000)	456	544
D <sub>c</sub> gr.cm <sup>-3</sup>	1.43	1.60
$\mu$ (MoK $\alpha$ ) mm <sup>-1</sup>	0.114	0.127
$((\sin \theta)/\lambda)_{\max}$ (Å <sup>-1</sup> )	0.55	0.55
Temperature (K)	298	298
No. measured reflc.	1614	1056
No. observed reflc.	1099	812
Criterion for observed	$ Fo  > 4\sigma(Fo)$	$ Fo  > 4\sigma(Fo)$
No. parameters	270	202
Weighting scheme	$1/\sigma^2(Fo)$	$1/\sigma^2(Fo)$
Max. and average $\Delta/\sigma$	0.018, 0.004	0.089, 0.011 (for H)
Max. and min. $\Delta\rho$ (e.Å <sup>-3</sup> )	0.38, -0.46	0.24, -0.26
S	1.23	1.92
R, $\omega R$ (%)	6.8, 6.7	3.6, 2.8

(E+Z)-2'-Deoxy-5,6-dihydro-2'-N-hydroxyimino-3',5'-O-(1'',1'',3'',3''-tetraisopropylidisiloxan-1'',3''-diyl)- $\beta$ -D-*erythro*-pentofuranosyluracil (2). A solution of 1 (5 g, 10 mmol) in CH<sub>2</sub>Cl<sub>2</sub> (200 mL) was treated at 0 °C with an excess of Garegg's reagent (CrO<sub>3</sub>, 9 g; pyridine, 15 mL; Ac<sub>2</sub>O, 9 mL). After completion of the reaction (TLC), the reaction mixture was filtered on a column of silica gel WOELM impregnated with AcOEt and the solvent

evaporated. The crude ketone was dissolved in a mixture of MeOH (100 mL) and pyridine (8 mL) and hydroxylamine chlorhydrate (2.1 g, 30.4 mmol) was added. After 2 h, the solvents were distilled, the residue extracted with AcOEt (3x50 mL) and the organic phase washed ( $H_2O$ , 100 mL) concentrated and submitted to column chromatography (1:2 AcOEt/ $CH_2Cl_2$ ) to give **2a** (0.71 g, 11.5%), **2b** (2.09 g, 34%) and **3** (0.63 g, 9.8%).

Properties of **2a** : mp 96.6-97.2 °C,  $R_F$  0.35 (1:4 AcOEt/hexane);  $[\alpha]_D^{25}$  -96.1° ( $c$  1.0,  $CHCl_3$ ); UV ( $CHCl_3$ ) 259 nm (10600); IR (KBr) 3250 (OH), 3110 (NH), 1710 and 1690  $cm^{-1}$  (C=O); MS ( $m/z$ , %) 112 (100, Ur + H), 69 (74), 120 (60), 146 (44), 135 (43), 105 (39), 175 (39), 289 (22), 163 (15), ...456 (11,  $M^+$  -  $CHMe_2$ ).

*Anal.* Calcd for  $C_{21}H_{37}N_3O_7Si_2$  (499.72): C, 50.48; H, 7.46; N, 8.41. Found: C, 50.32; H, 7.52; N, 8.19.

Properties of **2b** : mp 101.1-101.8 °C;  $R_F$  0.11 (1:4 AcOEt/ $CH_2Cl_2$ );  $[\alpha]_D^{21}$  -67° ( $c$  1.6,  $CHCl_3$ ); UV ( $CHCl_3$ ) 259 nm (10200); IR (KBr) 3300 (OH), 1720 and 1690  $cm^{-1}$  (C=O); MS ( $m/z$ , %) 112 (100, Ur + H), 69 (90), 317 (66), 456 (48,  $M^+$  -  $CHMe_2$ ), 329 (46), 120 (33), 344 (31), 147 (26), 135 (23), 175 (22).

*Anal.* calcd for  $C_{21}H_{37}N_3O_7Si_2$  (499.72): C, 50.48; H, 7.46; N, 8.41. Found: C, 50.33; H, 7.46; N, 8.32.

**(6S)-6,2'-Anhydro-5,6-dihydro-6-hydroxy-2'-N-hydroxyamino-3',5'-O-(1'',1'',3'',3'')-tetraisopropylidisiloxan-1''-3''-diyl)uridine (3)**. Obtained as described for the preparation of **2**. Mp 108.5-109.6 °C;  $R_F$  0.26 (1:10 MeOH/ $CH_2Cl_2$ );  $[\alpha]_D^{23}$  +6° ( $c$  1.0,  $CHCl_3$ ); UV ( $CHCl_3$ ) 240 nm (1300); IR (KBr) 3490 (OH), 3230 (NH), 1720 and 1690  $cm^{-1}$  (C=O, C=N). MS ( $m/z$ , %) 70 (100), 119 (97), 235 (86), 135 (85), 55 (78), 105 (68), 147 (62), 120 (57), 123 (53), ...474 (4,  $M^+$  -  $CHMe_2$ ).

*Anal.* Calcd for  $C_{21}H_{39}N_3O_8Si_2$  (517.73): C, 48.72; H, 7.59; N, 8.12. Found: C, 48.64; H, 7.68; N, 8.09.

**(6S)-6,2'-Anhydro-5,6-dihydro-6-hydroxy-2'-N-hydroxyamino-2'-O-methyl-3',5'-O-(1'',1'',3'',3'')-tetraisopropylidisiloxan-1'',3''-diyl)uridine (4)**. To a solution of **3** (0.91 g, 1.75 mmol) in MeOH (50 mL), HCl 3N in MeOH was added and the solution stirred at room temp for 15 h. The reaction mixture was brought to pH 6 (saturated aqueous  $NaHCO_3$ ), concentrated and submitted to a column chromatography (1:6 MeOH/ $CH_2Cl_2$ ) to give **4** (0.43 g, 94.4 %) as very hygroscopic crystals: mp 99.6-101 °C;  $R_F$  0.35 (1:6

MeOH/CH<sub>2</sub>Cl<sub>2</sub>); [ $\alpha$ ]<sub>D</sub><sup>26</sup> -48.3° (*c* 1.2, MeOH); UV (MeOH) 209 nm (5240); IR (KBr) 3380, 3240 (OH, NH), 1710, and 1690 cm<sup>-1</sup> (C=O); MS (*m/z*, %) 112 (100, Ur + H), 69 (52), 107 (16), 128 (10), 145 (6), 150 (6), 146 (4), 151 (3), 239 (0.3, M<sup>+</sup> - MeOH - H<sub>2</sub>O), 257 (0.2, M<sup>+</sup> - MeOH).

*Anal.* calcd for C<sub>10</sub>H<sub>15</sub>N<sub>3</sub>O<sub>7</sub> (289.25): C, 41.53; H, 5.23; N, 14.53. Found: C, 41.49; H, 5.61; N, 14.04.

**(6S)-3',5'-di-O-Acetyl-2'-N-acetoxymino-6,2'-anhydro-5,6-dihydro-6-hydroxyuridine (5).** A solution of compound 4 (200 mg, 0.69 mmol) in a mixture of pyridine (5 mL) and acetic anhydride (2 mL) was stirred at room temp for 10 h, then treated as usual. The analytical sample of 5 (127 mg, 41%) was obtained by column chromatography (1:1 AcOEt/hexane) followed by recrystallization (EtOH/H<sub>2</sub>O): mp 122.2-122.7 °C; [ $\alpha$ ]<sub>D</sub><sup>25</sup> -11.1° (*c* 0.7, CHCl<sub>3</sub>); UV (CHCl<sub>3</sub>) 240 nm (400); IR (KBr) 3260 (NH), 1790, 1770, 1740 and 1710 cm<sup>-1</sup> (C=O); MS (*m/z*, %) 373 (100, M<sup>+</sup> - AcOH), 331 (66), 112 (26), 113 (26), 299 (21), 198 (19), 356 (19), 341 (18), 230 (14).

*Anal.* calcd for C<sub>16</sub>H<sub>21</sub>N<sub>3</sub>O<sub>10</sub>·H<sub>2</sub>O (433.38); C, 44.34; H, 5.35; N, 9.70. Found: C, 43.99; H, 5.23; N, 9.58.

**(Z+E)-2'-Deoxy-2'-N-hydroxyimino-β-D-erythro-pentofuranosyluracil (6).** To a solution of 2 (0.5 g, 1 mmol) in methanol (40 mL), 2N HCl in MeOH (2 mL) was added and the mixture stirred at room temp for 38 h. The pH was brought to 6 (saturated aqueous NaHCO<sub>3</sub>) and the concentrated reaction mixture, submitted to a column chromatography (1:4 MeOH/CH<sub>2</sub>Cl<sub>2</sub>), gave a 3:7 mixture (193 mg, 75%) of the two geometrical isomers (6a and 6b) of 6 as a hygroscopic solid: mp 130-130.3 °C; *R*<sub>F</sub> 0.33 (6a) and 0.43 (6b) (1:4 MeOH/CH<sub>2</sub>Cl<sub>2</sub>); [ $\alpha$ ]<sub>D</sub><sup>22</sup> -43° (*c* 0.8, MeOH); UV (MeOH) 206 (9000) and 259 nm (6950); IR (KBr) 3380, 3240 (OH, NH) 1710, 1690 and 1660 cm<sup>-1</sup> (C=O, C=N); MS (*m/z*, %) 112 (100, Ur + H), 69 (66), 145 (21), 128 (7), 226 (7), 257 (3, M<sup>+</sup>), 239 (2, M<sup>+</sup> - H<sub>2</sub>O), 258 (1, M<sup>+</sup> + H).

*High resolution mass spectrum:* calcd for C<sub>9</sub>H<sub>11</sub>N<sub>3</sub>O<sub>6</sub> 257.0647. Found: 257.0672.

**(6S)-2'-deoxy-5,6-dihydro-6,2'-N-hydroxyimino-β-D-arabino-furanosyluracil (7).** To a solution of 2 (1.02 g, 2.04 mmol) in MeOH (50 mL) 3N HCl in MeOH (2 mL) was added and the solution stirred at room temp for 24 h to give 6 which was not isolated but reduced to 8 with BH<sub>3</sub>-pyridine (1 mL, 9.9 mmol) for 24 h at room temp. The reaction mixture was then

brought to pH 5 (saturated aqueous  $\text{NaHCO}_3$ ), concentrated to dryness, the residue washed with  $\text{CH}_2\text{Cl}_2$  (20 mL), then extracted with water (3x10 mL). The aqueous solution was concentrated to dryness, the residue dissolved in MeOH (30 mL) and reacted 15 h at room temp with veratraldehyde (0.5 g, 3.01 mmol). The reaction mixture concentrated gave, after column chromatography (1:10 MeOH/ $\text{CH}_2\text{Cl}_2$ ) a 3:7 *arabino/ribo* mixture of **10** (154 mg, 18.5 %) and **7** (0.4 g, 75%): mp 196.5-198.2 °C;  $R_F$  0.3 (1:4 MeOH/ $\text{CH}_2\text{Cl}_2$ );  $[\alpha]_D^{25} +130^\circ$  (c 1.0, MeOH); UV (MeOH) 204 nm (6400); IR (KBr) 3390, 3290, 3240 (OH, NH), 1720, and 1680  $\text{cm}^{-1}$  (C=O); MS ( $m/z$ , %) 113 (100), 152 (73), 70 (65), 84 (60), 130 (51), 169 (51), 81 (46), 56 (37), 97 (20) ...259 (3,  $M^+$ ).

*Anal.* calcd for  $\text{C}_9\text{H}_{13}\text{N}_3\text{O}_6$  (259.22): C, 41.70; H, 5.06; N, 16.21. Found: C, 41.67; H, 5.08; N, 16.18.

**(6S)-3',5'-di-O-Acetyl-6,2'-N-acetoxylimino-2'-deoxy-5,6-dihydro- $\beta$ -D-arabinofuranosyluracil (9).** A solution of **7** (0.16 g, 0.6 mmol) in a mixture of pyridine (10 mL) and acetic anhydride (5 mL) was stirred at room temp for 1 h, concentrated, then the solvents codistilled with toluene. Water (20 mL) was added to the residue and the mixture extracted with  $\text{CH}_2\text{Cl}_2$  (3x10 mL). The organic phases were dried ( $\text{Na}_2\text{SO}_4$ ), concentrated and submitted to a column chromatography (1:1 AcOEt/ $\text{CH}_2\text{Cl}_2$ ) which yielded **9** (152 mg, 64%); mp 73.0-73.9 °C;  $R_F$  0.25 (1:1 AcOEt/ $\text{CH}_2\text{Cl}_2$ );  $[\alpha]_D^{29} -43^\circ$  (c 0.6,  $\text{CHCl}_3$ ); UV (EtOH) 256 nm (1040); IR (KBr) 3240 (NH), 1760, 1730 and 1690  $\text{cm}^{-1}$  (C=O); MS ( $m/z$ , %) 81 (100), 113 (50, Ur + 2 H), 112 (43, Ur + H), 60 (38), 70 (34), 109 (28), 152 (21), 97 (20), 121 (13), ...325 (1,  $M^+$  - AcOH).

*Anal.* calcd for  $\text{C}_{15}\text{H}_{19}\text{N}_3\text{O}_9$  (385.33): C, 46.76; H, 4.97; N, 10.90. Found: C, 46.71; H, 5.05; N, 10.81.

**2'-Deoxy-2'-(3'',4''-dimethoxybenzylidenimino)- $\beta$ -D-arabino (and ribo)furanosyluracil  $N'$ -oxyde (10).** Obtained as described for the preparation of **7**: mp > 220 °C;  $R_F$  0.2 (1:10 MeOH/ $\text{CH}_2\text{Cl}_2$ ); UV (EtOH) 205 (19800), 231 (12200), 264 (12500), and 320 nm (13000); IR (KBr) 3400, 3230 (OH, NH), 1690 (C=O) and 1590  $\text{cm}^{-1}$  (C=N); MS ( $m/z$ , %) 112 (100, Ur + H), 69 (64), 165 (50), 77 (18), 261 (17), 231 (16), 151 (16), 259 (13), 247 (10), ...295 (4,  $M^+$  - UrH).

*Anal.* calcd for  $\text{C}_{18}\text{H}_{21}\text{N}_3\text{O}_8 \cdot \text{H}_2\text{O}$  (425.40): C, 50.82; H, 5.45; N, 9.88. Found: C, 50.59; H, 5.43; N, 9.47.



## ACKNOWLEDGEMENTS

The authors thank the Swiss National Science Foundation for grants (20-26460.89 and 20-31259.91), Prof. M. Geoffroy for the EPR spectra, Dr N. Dolatshahi for the biological testing, and Dr H. Eder for the elementary analyses.

## REFERENCES

1. Tronchet, J. M. J. in "Bioactive Spin Labels", R. Zhdanov Ed. Springer Verlag Berlin (1992) pp 355-387.
2. Tronchet, J. M. J.; Benhamza, R.; Dolatshahi, N.; Geoffroy, M.; Türlér, H., *Nucleosides & Nucleotides*, **1988**, 7, 249.
3. Jenks, W. P., *J. Am. Chem. Soc.*, **1958**, 80, 4581; Klopman, G.; Tsuda, K.; Louis, J. B.; Davis, R. E., *Tetrahedron Lett.*, **1970**, 26, 4549.
4. Doerr, I. L.; Cushley, R. J.; Fox, J. J., *J. Org. Chem.*, **1968**, 33, 1952; Doerr, I. L.; Fox, J. J., *J. Am. Chem. Soc.*, **1967**, 89, 1760; Shibuya, S.; Ueda, T., *Chem. Pharm. Bull.*, **1980**, 28, 939.
5. Tronchet, J. M. J.; Benhamza, R.; Bernardinelli, G., Geoffroy, M., *Tetrahedron Lett.* **1990**, 31, 531 .
6. Robins, M. J.; Wilson, J. S.; Sawyer, L.; James, M. N. G., *Can. J. Chem.*, **1983**, 61, 1911; Markiewicz, W. T.; Padyuhova, N. S.; Samek, Z.; Smrt, J. *Collect. Czech. Commun.*, **1980**, 45, 1860.
7. Garegg, P. J.; Samuelsson, B. *Carbohydr. Res.*, **1978**, 67, 267; Garegg, P. J.; Maron, L., *Acta Chem. Scand.*, **1979**, B33, 453.
8. Sundaralingam, M., *J. Amer Chem. Soc.*, **1971**, 93, 6644.
9. Nardelli, M., *Acta Crystallogr.*, **1983**, C39, 1141.
10. Cremer, D.; Pople, J. A., *J. Am. Chem. Soc.*, **1975**, 97, 1354.
11. Sundaralingam, M., *Biopolymers*, **1969**, 7, 821.
12. Saenger, W.; Scheit, K. H. *J. Mol. Biol.*, **1970**, 50, 153.
13. Tronchet, J. M. J.; Bizzozero, N.; Koufaki, M.; Habashi, F.; Geoffroy, M., *J. Chem. Res.*, **1989**, (S) 334, (M) 2601.
14. Rassat, A.; Lemaire, H., *J. Chim. Phys.*, **1964**, 61, 1576.
15. Meyer zu Reckendorf, W., *Chem. Ber.*, **1963**, 96, 2019.
16. Loew, B.; Goodman, M. M., *Chem. Ind. (London)*, **1967**, 2026.
17. Still, W. C.; Kahn, M.; Mitra, A., *J. Org. Chem.*, **1978**, 43, 2923.

18. Barbalat-Rey, F.; Lichtle, P., unpublished results.
19. Main, P.; Fiske, S. J.; Hull, S. E.; Lessinger, L.; Germain, G.; Declercq, J.-P.; Woolfson, M. M. (1987). *A System of Computer Programs for the Automatic Solution of Crystal Structures from X-Ray Diffraction Data*. Univs. of York, England, and Louvain-la-Neuve, Belgium.
20. Hall, S. R.; Stewart, J. M. *Eds XTAL2.5 User's Manual*, Universities of Western Australia and Maryland, (1987).

Received 5/21/92

Accepted 9/24/92